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Product Sheet

H_IL22 Reporter 293 Cell Line

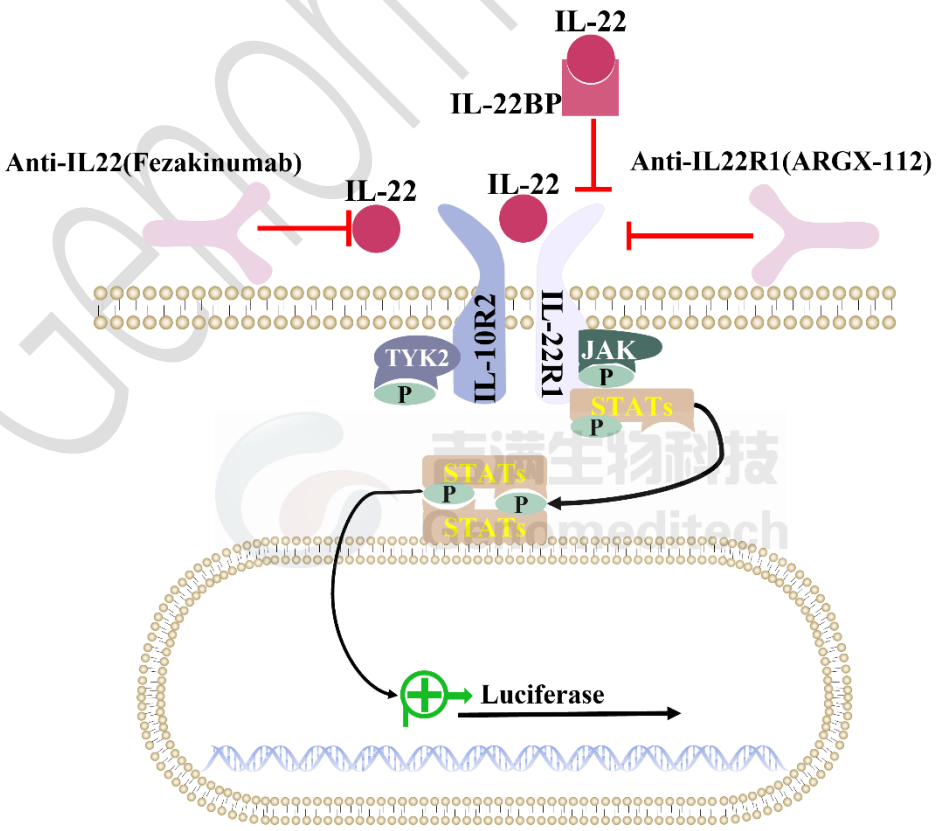
Catalog number: GM-C19682

Version 3.3.1.241217

IL-22 is a cytokine belonging to the IL-10 family, primarily produced by activated T cells and innate lymphoid cells (ILCs). It plays an important role in maintaining tissue homeostasis and defending against infections, particularly in the protection of epithelial cells. IL-22 exerts its biological effects by binding to its receptor composed of IL-22R1 and IL-10R2 heterodimers, activating the downstream JAK1-STAT3 signaling pathway.

By binding to the heterodimeric receptor formed by IL-22R1 and IL-10R2, IL-22 activates the JAK1-STAT3 signaling pathway, primarily activating STAT3, but also capable of activating STAT1 and STAT5. Additionally, IL-22 can trigger the NF-κB, MAPK, and PI3K-Akt-mTOR signaling pathways.

H_IL22 Reporter 293 Cell Line is a clonal stable HEK-293 cell line constructed using lentiviral technology, constitutive expression of the IL-22R1 gene and IL-10R2 gene, along with signal-dependent expression of a luciferase reporter gene. When IL-22 binds to IL-22R1 and IL-10R2 heterodimers, it activates downstream signaling pathways, leading to the expression of luciferase. Blockade antibodies can inhibit this signal transmission. The luciferase activity measurement indicates the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to IL-22.



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Specifications

Quantity	5E6 Cells per vial,1 mL
Product Format	1 vial of frozen cells
Shipping	Shipped on dry ice
Storage Conditions	Liquid nitrogen immediately upon receipt
Recovery Medium	DMEM+10% FBS+1% P.S
Growth medium	DMEM+10% FBS+1% P.S+4 μg/mL Blasticidin+400 μg/mL G418+125 μg/mL Hygromycin+0.75 μg/mL Puromycin
Note	None
Freezing Medium	90% FBS+10% DMSO
Growth properties	Adherent
Growth Conditions	37°C, 5% CO_2
Mycoplasma Testing	The cell line has been screened to confirm the absence of Mycoplasma species.
Safety considerations	Biosafety Level 2
Note	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

Materials

Reagent	Manufacturer/Catalogue No.
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/GM-040404
G418	Genomeditech/GM-040402
Hygromycin	Genomeditech/GM-040403
Puromycin	Genomeditech/GM-040401
Recombinant Human IL-22 Protein	Sino Biological/13059-HNAE
Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112)	Genomeditech/GM-46382AB
Anti-H_IL-22 hIgG1 Antibody(Fezakinumab)	Genomeditech/GM-46509AB
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/GM-040503

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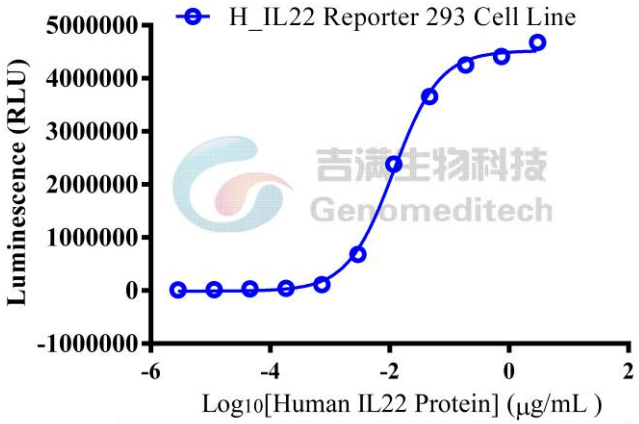
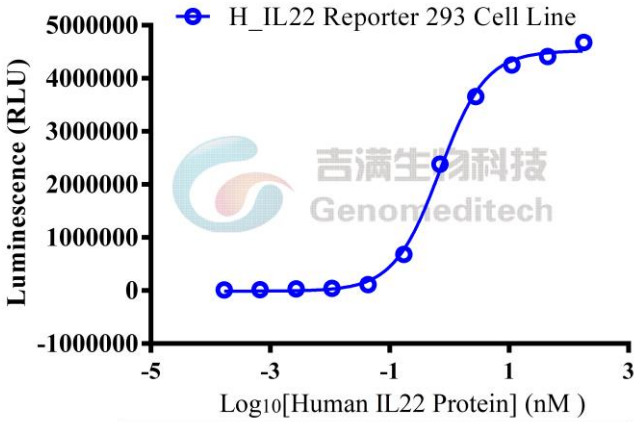
Figures



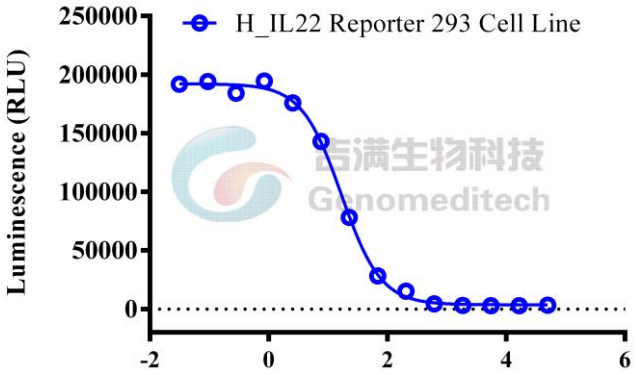
	H_IL22 Reporter 293 Cell Line
EC50	0.01168

Figure 1 | Response to Recombinant Human IL-22 Protein. The H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) in assay buffer (DMEM + 1% FBS + 1% P.S) for 16 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The maximum induction fold was approximately [440.9]. Data are shown by drug mass concentration.



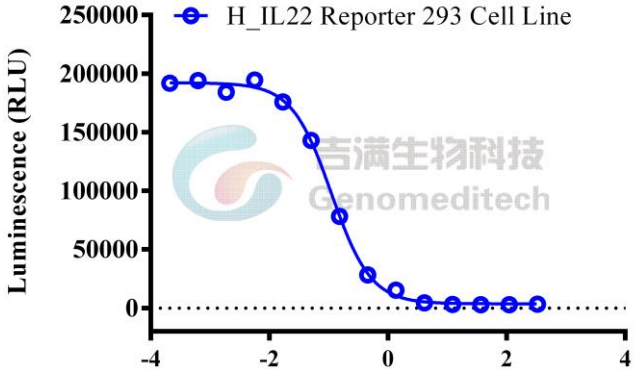
	H_IL22 Reporter 293 Cell Line
EC50	0.6911

Figure 2 | Response to Recombinant Human IL-22 Protein. The H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) in assay buffer (DMEM + 1% FBS + 1% P.S) for 16 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The maximum induction fold was approximately [440.9]. Data are shown by drug molar concentration.



	H_IL22 Reporter 293 Cell Line
IC50	16.72

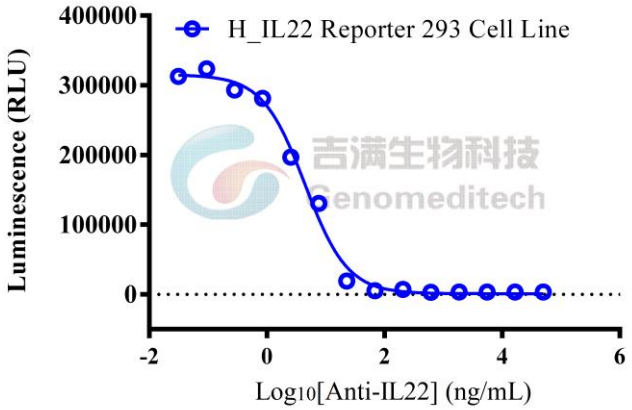
Figure 3 | Response to Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112). Serial dilutions of the Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112) (Cat. GM-46382AB) was incubated with 1.5E4 cells/well of the H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). Subsequently, the Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) at a concentration of 1 ng/well was added, and the coculture proceeded for an additional 15 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The results indicated maximum blocking folds of approximately [59.0]. Data are shown by drug mass concentration.



	H_IL22 Reporter 293 Cell Line
IC50	0.1114

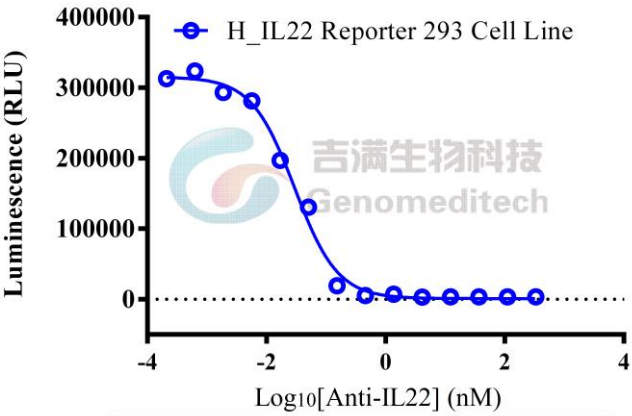
Figure 4 | Response to Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112). Serial dilutions of the Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112) (Cat. GM-46382AB) was incubated with 1.5E4 cells/well of the H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). Subsequently, the Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) at a concentration of 1 ng/well was added, and the coculture proceeded for an additional 15 hours. Firefly luciferase activity is then measured using the GMOne-

Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The results indicated maximum blocking folds of approximately [59.0]. Data are shown by drug molar concentration.



	H_IL22 Reporter 293 Cell Line
IC50	4.439

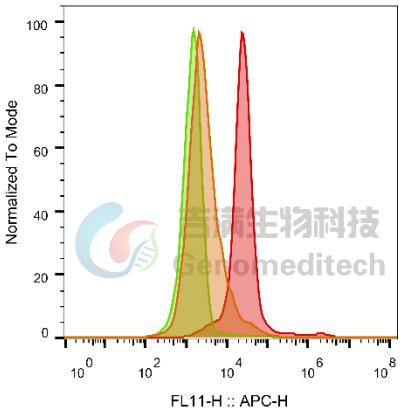
Figure 5 | Response to Anti-H_IL-22 hIgG1 Antibody(Fezakinumab). Serial dilutions of Anti-H_IL-22 hIgG1 Antibody(Fezakinumab) (Cat. GM-46509AB) was incubated with 1 ng/well of Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). After pre-incubation, add the mixture to the H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) at a density of 1.5E4 cells/well in a 96-well format, and incubate for 15 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The results indicated maximum blocking folds of approximately [95.5]. Data are shown by drug mass concentration.



	H_IL22 Reporter 293 Cell Line
IC50	0.02959

Figure 6 | Response to Anti-H_IL-22 hIgG1 Antibody(Fezakinumab). Serial dilutions of Anti-H_IL-22 hIgG1 Antibody(Fezakinumab) (Cat. GM-46509AB) was incubated with 1 ng/well of Recombinant Human IL-22 Protein (SinoBiological/13059-HNAE) for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). After pre-incubation, add the

mixture to the H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) at a density of 1.5E4 cells/well in a 96-well format, and incubate for 15 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The results indicated maximum blocking folds of approximately [95.5]. Data are shown by drug molar concentration.



SampleID	Geometric Mean : FL11-H
HEK-293 anti-IL22R1+APC-2nd Ab	2793
H_IL22 Reporter 293 H_IgG+APC-2nd Ab	1462
H_IL22 Reporter 293 anti-IL22R1+APC-2nd Ab	25673

Figure 7 | H_IL22 Reporter 293 Cell Line (Cat. GM-C19682) was determined by flow cytometry using Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112) (Cat. GM-46382AB).

Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).

b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.

c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.

d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.

e) Incubate the culture at 37°C in a suitable incubator. A 5% CO_2 in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

a) Centrifuge at 176 x g for 3 minutes to collect cells.

b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 5E6 cells/mL.

c) Aliquot 1 mL into each vial.

d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: DMEM+10% FBS+1% P.S+4 μg/mL Blasticidin+400 μg/mL G418+125 μg/mL Hygromycin+0.75 μg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

a) Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.

b) Remove and discard culture medium.

c) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.

d) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at 37°C).

e) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.

f) Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.

h) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.

b) Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

Related Products

IL-22	
Anti-H_IL-22 hIgG1 Antibody(Fezakinumab)	Anti-H_IL-22R1 hIgG1 Antibody(ARGX-112)

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